

Genetic and Environmental Influences on Lipids, Lipoproteins, and Apolipoproteins

Effects of Menopause

Rita P.S. Middelberg, Tim D. Spector, Ramasamyiyer Swaminathan, Harold Snieder

Objective—Levels of lipids and (apo)lipoproteins are known to increase after menopause, but it is unknown whether the genetic and environmental variability alters or whether lipids and (apo)lipoproteins are influenced by different genes before and after menopause.

Methods and Results—We studied 453 monozygotic and 1280 dizygotic pairs of female white twins recruited from the St. Thomas' UK Adult Twin Registry and measured total cholesterol, low density lipoprotein (LDL), high density lipoprotein (HDL), triglycerides, lipoprotein(a) [Lp(a)], apolipoprotein A1 (apoA1), and apolipoprotein B (apoB). Variance components software was used to estimate genetic and environmental influences on serum lipid levels in premenopausal and postmenopausal women. Total variance was higher for triglycerides, HDL, and apoB after menopause. Postmenopausal women showed larger genetic variance for most lipids, apart from apoB and Lp(a). In premenopausal females, total cholesterol, LDL, HDL, apoA1, and apoB all showed an influence of the shared environment (22% to 34%), which, after menopause, decreased in HDL and completely disappeared in total cholesterol, LDL, and apoA1. Only for Lp(a), with a high heritability of 87%, did the same model fit premenopausal and postmenopausal women. Generally, there was no indication that different genes influence lipids before and after menopause.

Conclusions—These findings imply that genetic studies of lipids can pool results from premenopausal and postmenopausal women and that family-based interventions, such as changes in diet, are more likely to succeed in younger women, in whom the environmental influences are greater. (*Arterioscler Thromb Vasc Biol.* 2002;22:1142-1147.)

Key Words: lipids ■ genetics ■ twin study ■ menopause

Postmenopausal women are 2 to 3 times more likely to suffer from coronary heart disease than are premenopausal women.^{1,2} Part of this increased risk is likely to be due to adverse changes in lipids and (apo)lipoproteins after menopause. Levels of total cholesterol, LDL, triglycerides, and apoB have been reported to increase,^{3,4} probably (at least partly) because of a reduction in LDL receptor activity in response to the gradual decline in blood estrogen levels in the perimenopausal years.^{5,6} Levels of lipids, lipoproteins, and apolipoproteins, which have been established as important predictors of atherosclerotic coronary disease,⁷ vary considerably over a lifespan, and their phenotypic variance generally shows an increase with age.⁸⁻¹¹ Such an increase in lipid and (apo)lipoprotein variance may be due to interindividual variation in the rise of lipid levels over time, and several studies now indicate that different genetic and/or environmental factors are likely to be involved at different ages.^{10,12,13} Superimposed on the global change across the whole lifespan, 4 specific time periods are associated with

more dramatic changes. Menopause is one of these; the others are old age, adolescence, and the first years after birth.

We recently reviewed twin studies of lipids and (apo)lipoproteins and showed that most studies report heritability (h^2) estimates between 40% and 80% for total cholesterol, LDL, HDL, triglycerides, apoA1, and apoB, with Lp(a) h^2 values of $\approx 90\%$.¹¹ Only one study, which reported on HDL alone, focused on the effect of menopause and found that postmenopausal monozygotic (MZ) twins were more similar ($r=0.79$) than premenopausal MZ twins ($r=0.61$), whereas dizygotic (DZ) similarity did not change ($r=0.31$ and $r=0.32$, respectively).¹⁴ These results suggest that genetic mechanisms that affect individual variation in HDL level may differ in premenopausal and postmenopausal women. For the other lipids and (apo)lipoproteins, it is unknown whether an increase in phenotypic variance after menopause is due to genes, environment, or both or whether they may be influenced by different genes before and after menopause.

Received January 12, 2002; revision accepted April 23, 2002.

From the Twin Research and Genetic Epidemiology Unit (R.P.S.M., T.D.S., H.S.) and the Department of Chemical Pathology (R.S.), St. Thomas' Hospital, London, UK; Gemini Genomics (R.P.S.M.), Cambridge, UK; and the Georgia Prevention Institute (H.S.), Medical College of Georgia, Augusta.

Reprint requests to Dr Harold Snieder, Georgia Prevention Institute, Medical College of Georgia, Building HS-1640, Augusta, GA 30912-3700. E-mail hsnieder@mcg.edu

© 2002 American Heart Association, Inc.

Arterioscler Thromb Vasc Biol. is available at <http://www.atvbaha.org>

DOI: 10.1161/01.ATV.0000022889.85440.79

Therefore, the aim of this classic twin study was to quantify and compare the genetic and environmental sources of individual differences in lipids and (apo)lipoproteins in premenopausal and postmenopausal women. A secondary aim was to determine whether these traits are influenced to some degree by different genes before and after menopause, which may have an impact on the design of gene discovery studies. Quantitative genetic modeling techniques were used to analyze lipid data in a large sample of unselected female adult twins across a broad age range.

Methods

Subjects

The study cohort was composed of 1733 (453 MZ and 1280 DZ) female white twin pairs aged 18 to 79 years from the St. Thomas' UK Adult Twin Registry. Twins from the registry were ascertained from the general population through national media campaigns in the United Kingdom.¹⁵ Participating twins were unaware of the specific hypotheses tested, and informed consent was obtained from all subjects. The study was approved by the St. Thomas' Hospital Research Ethics Committee.

Measures

Subjects were interviewed and studied by trained research nurses. Zygosity was determined by standardized questionnaire, and DNA fingerprinting was used for confirmation.¹⁶ Information on (gynecological) medical history, medication use, fasting duration, lifestyle, and demographic variables was obtained by a standardized nurse-administered questionnaire. Postmenopausal status was defined as the cessation of menstruation for at least 12 months.^{17,18} Fasting status was coded as either yes (fasting ≥ 8 hours) or no (fasting < 8 hours). Height was measured to the nearest 0.5 cm by using a wall-mounted stadiometer. Weight (light clothing only) was measured to the nearest 0.1 kg by using digital scales. Body mass index was calculated as weight/height².

A venous blood sample was taken in the early morning after an overnight fast. For both twins of each pair, blood was taken ≤ 5 minutes apart. Serum samples were stored at -40°C until analysis. Levels of all lipids were measured by using a Cobas Fara machine (Roche Diagnostics). Total cholesterol, HDL, and triglycerides were determined by a colorimetric enzymatic method. HDL cholesterol was determined after precipitation of larger particles (chylomicron, VLDL, and LDL) by magnesium and dextran sulfate. ApoA1, apoB, and Lp(a) were determined by an immunoturbidometric method. LDL levels were estimated by using the Friedewald equation.¹⁹ This formula was applied only if the triglyceride concentration of subjects did not exceed 4.52 mmol/L. Four subjects on lipid-lowering

medication and 17 subjects with extremely high (or low) lipid values (>4 SD above or below the mean) were excluded from the analyses.

Statistical Analysis

Background for Twin Analysis

The classic twin study makes use of the fact that MZ twins share identical genotypes, whereas DZ twins are no more alike genetically than siblings, sharing, on average, 50% of their segregating genes. If MZ twins show a larger resemblance for a specific trait than do DZ twins, this is attributed to genetic factors. Genetic model fitting is based on the comparison of the variance-covariance matrices in MZ and DZ twin pairs and allows separation of the observed phenotypic variance into its genetic and environmental components: additive genetic variance, dominant genetic variance, shared (or common) environmental variance, and specific (or unique) environmental variance, which also contains measurement error. The term h^2 can be defined as the ratio of additive genetic variance to total phenotypic variance.²⁰

Analytical Approach

The distributions of total cholesterol, triglycerides, LDL, and Lp(a) were skewed and transformed by natural logarithm to normalize distributions before analysis. Preliminary data analysis was performed, and intraclass correlations were calculated by using STATA.²¹

Each of the lipids and (apo)lipoproteins was adjusted for the effect of age, fasting, menopausal status, and use of hormone replacement therapy (HRT) by multiple regression, and trait residuals were saved. All model fitting was performed on these residuals.²²

The aims of our analysis were, first, to establish the genetic influence overall by estimating the genetic and environmental influences on all variables for the entire group. Second, we investigated the effect of menopause on changes in (1) total trait variance and (2) genetic and environmental sources of variance. We specified a path model in which the total sample was subdivided into 6 zygosity-by-menopause groups (ie, MZ pre/pre, DZ pre/pre, MZ pre/post, DZ pre/post, MZ post/post, and DZ post/post, where pre/pre indicates twins that are both premenopausal, pre/post indicates 1 premenopausal and 1 postmenopausal twin, and post/post indicates twins that are both postmenopausal) to estimate variance components in premenopausal and postmenopausal women separately.²³ Using the principle of parsimony, we subsequently homed in on the best fitting model, ie, the simplest model (in terms of number of parameters estimated) that still described the data well. If traits showed equal total variance and the same model in premenopausal and postmenopausal women, we tested whether variance components could be set equal for premenopausal and postmenopausal women.

Finally, we examined whether in menopause-discordant pairs the correlation between the latent genetic factors (r_g) was lower than the

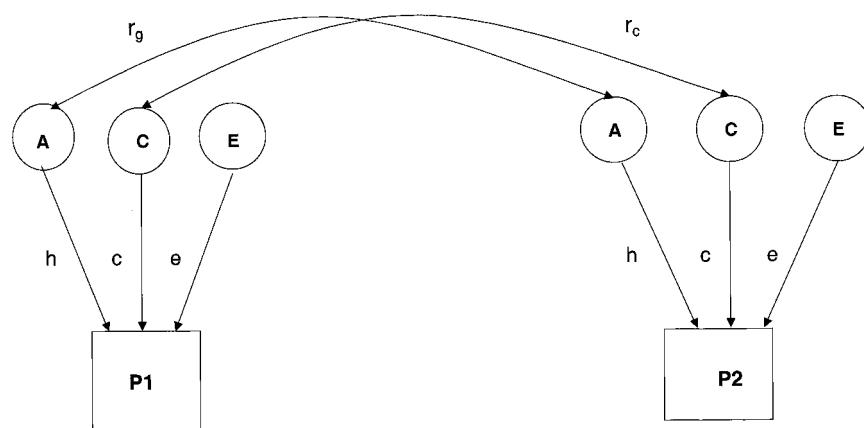


Figure 1. Path diagram for twin model. Observed variables for twin 1 and twin 2 are shown in the squares. Latent variables (or factors) are shown in circles. A single-headed arrow indicates a direct influence of one variable on another; its value is represented by a path coefficient. Double-headed arrows indicate a correlation without any assumed direct relationship. A indicates additive genetic factor; C, common environmental factor; E, unique environmental factor; h, additive genetic factor loading; c, common environmental factor loading; e, unique environmental factor loading; P1 (P2), phenotypic value of twin 1 (twin 2); r_g , genetic correlation (1 for MZ and 0.5 for DZ twins); and r_c , common environmental correlation (1 for MZ and DZ twins). r_g and r_c can be estimated in menopausal-discordant pairs.

TABLE 1. General Characteristics of Study Population

	MZ	DZ
No. of pairs	453	1280
Age, y	50.56 (13.61)	47.07 (11.68)
Weight, kg	64.41 (10.31)	66.06 (12.70)
Height, cm	162.35 (6.18)	162.51 (6.11)
BMI, kg/m ²	24.44 (4.02)	25.01 (4.68)
Postmenopausal, %	61.92	46.95
Current smokers, %	19.07	21.98
Current HRT user, %	14.09	18.20
Current OCP user, %	12.37	10.93
Hysterectomy, %	16.26	15.68
Bilateral oophorectomy, %	4.03	4.43

Mean (SD) is shown, unless stated otherwise. BMI indicates body mass index; OCP, oral contraceptive pill.

normal expected correlation of 1 in MZ pairs and 0.5 in DZ pairs, which would indicate that (partly) different genes influence the lipids and (apo)lipoproteins in premenopausal and postmenopausal women. Alternatively, it was tested whether discordant twins share their environment to a lesser extent than concordant pairs, ie, whether the shared environmental correlation (r_e) is lower than the expected correlation of 1 in MZ and DZ pairs (Figure 1).²³

Model-Fitting Procedure

Models were fitted to the raw data by using normal theory maximum likelihood.²⁴ This method allowed us to use the information provided by unpaired observations, which contribute to the estimation of variance (but not covariance).

The significance of additive genetic variance, common environmental variance, and dominant genetic variance was tested by removing each sequentially in specific submodels. Variance components were dropped from the model if they did not give a significant contribution (ie, $P > 0.05$). Submodels were compared with the full model by hierarchic χ^2 tests. In comparing non-nested models, the best model was chosen on the basis of the lowest value of Akaike's information criterion ($\chi^2 - 2 df$), which reflects the best balance between goodness of fit and parsimony. Estimates of variance components and their 95% CIs were obtained from the best fitting model. All quantitative genetic model fitting was carried out by statistical modeling using Mx.²⁵

Results

Results for the Entire Group

The data in Table 1 show the general characteristics of MZ and DZ twins for the entire study population of 1733 twin pairs. Comparison of the groups revealed that MZ pairs were, on average, 3.5 years older than DZ pairs at the time of ascertainment and that a larger proportion were, consequently, postmenopausal. All other charac-

TABLE 2. Mean (SD) of Lipid Measurements for MZ and DZ Twins

	MZ	DZ
Total cholesterol, mmol/L	5.75 (1.32)	5.52 (1.21)
LDL, mmol/L	3.64 (1.22)	3.44 (1.10)
HDL, mmol/L	1.55 (0.37)	1.54 (0.39)
Triglycerides, mmol/L	1.37 (0.89)	1.23 (0.76)
ApoA1, g/L	1.74 (0.35)	1.67 (0.34)
ApoB, g/L	1.25 (0.38)	1.14 (0.35)
Lp(a), mg/dL	28.75 (32.33)	29.03 (32.12)

TABLE 3. Intraclass Correlation Coefficients (Number of Complete Twin Pairs) for MZ and DZ Twins by Menopausal Status

	Menopausal Status			
	Pre/Pre	Pre/Post	Post/Post	Total
Total cholesterol				
MZ	0.59 (161)	0.18 (18)	0.63 (263)	0.61 (442)
DZ	0.42 (587)	0.37 (148)	0.33 (510)	0.38 (1245)
LDL				
MZ	0.57 (142)	0.62 (13)	0.71 (249)	0.66 (404)
DZ	0.46 (559)	0.28 (141)	0.33 (479)	0.39 (1179)
HDL				
MZ	0.66 (162)	0.45 (17)	0.63 (267)	0.64 (446)
DZ	0.50 (590)	0.30 (150)	0.46 (509)	0.46 (1249)
Triglycerides				
MZ	0.59 (144)	0.88 (14)	0.64 (258)	0.63 (416)
DZ	0.29 (568)	0.20 (145)	0.40 (494)	0.33 (1207)
ApoA1				
MZ	0.62 (147)	0.58 (14)	0.51 (238)	0.55 (399)
DZ	0.45 (551)	0.17 (137)	0.31 (476)	0.36 (1164)
ApoB				
MZ	0.76 (148)	0.58(14)	0.63 (240)	0.68 (402)
DZ	0.45 (535)	0.50 (136)	0.43 (474)	0.45 (1145)
Lp(a)				
MZ	0.85 (147)	0.93 (14)	0.88 (240)	0.87 (401)
DZ	0.50 (541)	0.34 (138)	0.47 (477)	0.46 (1156)

teristics were very similar for MZ and DZ twins. The mean values for all lipids, lipoproteins, and apolipoproteins were similar for both groups and are shown in Table 2.

The intraclass correlations for the entire group of MZ and DZ twins for the different lipid variables are presented in the last column of Table 3. For each of the measures, MZ correlations were greater than DZ correlations, implying an important genetic influence, which was subsequently confirmed by model fitting to the entire group (Table 4). Dominant genetic effects did not contribute significantly to the explanation of the data for any of the lipid variables; ie, dominant genetic effects could be dropped from the model without a significant worsening of the fit. A model specifying additive genetic, common environmental, and unique environmental variance components gave the

TABLE 4. Variance Component Estimates of Lipid Traits of Best Models Fitted to the Entire Twin Sample

	h^2 (95% CI)	c^2 (95% CI)	e^2 (95% CI)
Total cholesterol	0.42 (0.28, 0.56)	0.17 (0.07, 0.28)	0.40 (0.35, 0.46)
LDL	0.46 (0.32, 0.59)	0.17 (0.07, 0.28)	0.37 (0.32, 0.42)
HDL	0.42 (0.30, 0.54)	0.24 (0.15, 0.33)	0.34 (0.29, 0.39)
Triglycerides	0.63 (0.58, 0.67)	–	0.37 (0.33, 0.42)
ApoA1	0.36 (0.20, 0.51)	0.18 (0.07, 0.30)	0.45 (0.40, 0.52)
ApoB	0.39 (0.26, 0.51)	0.26 (0.16, 0.36)	0.35 (0.30, 0.40)
Lp(a)	0.87 (0.85, 0.89)	–	0.13 (0.11, 0.15)

h^2 indicates heritability; c^2 , shared environmental variance component, e^2 , unique environmental variance component.

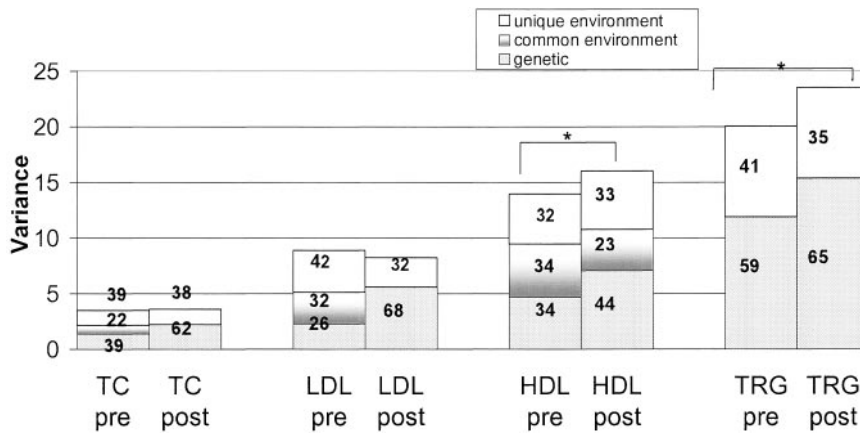


Figure 2. Total variance and standardized variance components of best fitting models for lipids and lipoproteins in premenopausal and postmenopausal women. Standardized variance components (percentage of total variance) are shown in each bar. TC indicates total cholesterol; TRG, triglycerides. * $P < 0.05$ for significant differences in total variance by menopausal status.

most parsimonious explanation of the data for all of the variables, with the exception of Lp(a) and triglycerides, for which shared environmental effects did not contribute significantly to the variation. Values of h^2 ranged from 0.36 for apoA1 to 0.87 for Lp(a) (Table 4).

Effect of Menopause

No significant differences for mean lipid values were found between premenopausal and postmenopausal twins within (age-identical) menopause-discordant twin pairs (data not shown). However, multiple regression using all the twin data showed significantly higher mean levels of total cholesterol ($P=0.006$), triglycerides ($P=0.002$), and apoB ($P=0.005$) in postmenopausal women after adjustment for age, fasting status, and HRT use.

Intraclass correlations for each of the 6 zygosity-by-menopause groups are shown in Table 3. Total variance was higher after menopause for HDL, triglycerides, and apoB and did not differ significantly between premenopausal and postmenopausal women for the other lipids (Figures 2 and 3). Genetic and environmental variance components estimates for premenopausal and postmenopausal women of the best fitting models are presented in Table 5 and Figures 2 and 3. Most lipids showed larger genetic variance in postmenopausal women, apart from apoB and Lp(a), for which the (nonstandardized) genetic variance remained stable. In premenopausal women, total cholesterol, LDL, HDL, apoA1, and apoB all showed an influence of the shared environment (22% to 34%), which, after menopause, decreased in HDL and completely disappeared in total cholesterol, LDL, and

apoA1. In the best fitting model for Lp(a), genetic and environmental variance components were identical for premenopausal and postmenopausal women ($h^2=87\%$).

Different Genes or Shared Environment Before and After Menopause

With the exception of HDL, there was no evidence that lipids or apolipoproteins are influenced by different genes or that the extent of environmental sharing differs before and after menopause. For most lipid variables, the genetic correlations (r_g values) were estimated at or very close to 1 and 0.5 for menopause-discordant MZ and DZ twins, respectively. The shared environmental correlations (r_c values) were estimated very close to 1 for both zygositys. For HDL, a model that specified less environmental sharing between discordant pairs ($r_c=0.28$) fitted slightly better than a model specifying different genetic influences before and after menopause ($r_g=0.13$), but variance components estimates were virtually identical for the 2 models. Both models were significantly better than a simpler model in which r_c and r_g were fixed to their usual values.

Discussion

The present study investigated whether the magnitude of genetic or environmental influences on lipids and (apo)lipoproteins changes with menopause and whether lipids and (apo)lipoproteins may be influenced by different genes before and after menopause. We observed that genetic variance in postmenopausal women was larger for most lipids and (apo)lipoproteins, except for apoB and Lp(a), which showed no change. Shared

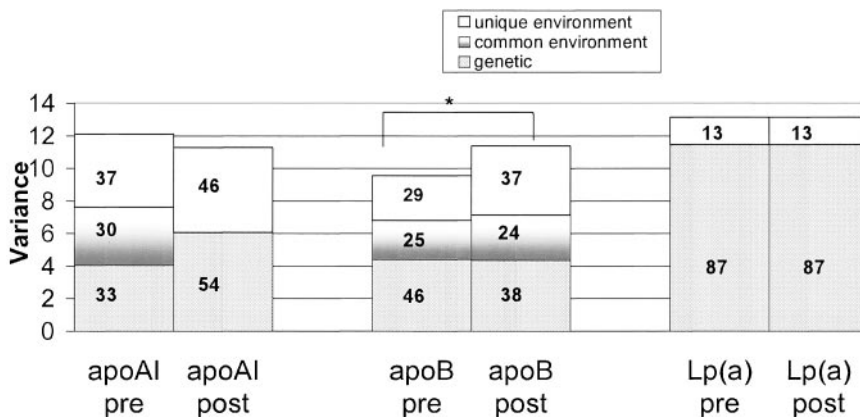


Figure 3. Total variance and standardized variance components of best fitting models for apolipoproteins and Lp(a) in premenopausal and postmenopausal women. Standardized variance components (percentage of total variance) are shown in each bar. Unit of variance for Lp(a) is 1/10 the original unit. * $P < 0.05$ for significant differences in total variance by menopausal status.

TABLE 5. Genetic and Environmental Variance Component Estimates of Best Fitting Models for Pre- and Postmenopausal Women

	Pre			Post		
	h ² (95% CI)	c ² (95% CI)	e ² (95% CI)	h ² (95% CI)	c ² (95% CI)	e ² (95% CI)
Total Cholesterol	0.39 (0.22, 0.57)	0.22 (0.08, 0.35)	0.39 (0.32, 0.47)	0.62 (0.56, 0.68)	–	0.38 (0.32, 0.44)
LDL	0.26 (0.09, 0.43)	0.32 (0.19, 0.44)	0.42 (0.35, 0.51)	0.68 (0.62, 0.73)	–	0.32 (0.27, 0.38)
HDL	0.34 (0.15, 0.50)	0.34 (0.21, 0.47)	0.32 (0.26, 0.40)	0.44 (0.27, 0.61)	0.23 (0.09, 0.36)	0.33 (0.27, 0.40)
Triglycerides	0.59 (0.50, 0.67)	–	0.41 (0.33, 0.50)	0.65 (0.59, 0.70)	–	0.35 (0.30, 0.41)
ApoA1	0.33 (0.16, 0.50)	0.30 (0.17, 0.42)	0.37 (0.30, 0.46)	0.54 (0.46, 0.61)	–	0.46 (0.39, 0.54)
ApoB	0.46 (0.28, 0.63)	0.25 (0.11, 0.39)	0.29 (0.23, 0.36)	0.38 (0.19, 0.56)	0.24 (0.09, 0.39)	0.37 (0.31, 0.45)
Lp(a)	0.87 (0.85, 0.89)	–	0.13 (0.11, 0.50)	0.87 (0.85, 0.89)	–	0.13 (0.11, 0.50)

environmental influences tended to be smaller or even disappear after menopause. No evidence was found indicating that lipids are influenced by different genes before and after menopause.

A recent review of twin studies of lipids and (apo)lipoproteins¹¹ showed that most studies report h² estimates between 40% and 80% for total cholesterol, LDL, HDL, and triglycerides. Estimates of h² for apoA1 and apoB were roughly within the same range as for lipid levels. Lp(a) h² values were ≈90%. None of the studies reviewed found much support for an important influence of family environment on the variance of the apolipoprotein and lipid levels.

The major difference between this and former twin studies is our finding that shared environment does contribute significantly to most lipid variables. Only for triglycerides and Lp(a) was no such influence detected. For the other lipids and apolipoproteins, between 17% and 26% of the total variance was explained by environmental factors common to both twins in the overall study sample. Premenopausal women showed even higher estimates of the shared environment, explaining between 22% and 34% of the variance. As might be expected, the influence of shared (familial) experiences on lipid levels wears off in later life. After menopause, shared environment decreased in HDL and disappeared completely in total cholesterol, LDL, and apoA1.

Because of the fact that the familial resemblance could be explained by sharing of environment in addition to sharing of genes, overall h² estimates (ie, ignoring menopause dependency) in the present study were slightly lower than reported in previous twin studies and ranged from 0.36 for apoA1 to 0.87 for Lp(a).

Hopper²⁶ has convincingly argued that most twin studies simply lack the power to detect moderate size influences of a common environment. The present study is the largest twin study to date investigating the relative contribution of genes and environment on lipids and apolipoproteins and is more than twice as large as the second largest twin study, which was published recently.²⁷ Thus, our results confirm that large sample sizes are needed to detect moderate influences of shared environment in twin studies.

A recent study of the Swedish Twin Registry²⁷ evaluated the effect of aging on genetic and environmental variation in lipid and apolipoprotein levels. Their results showed that increases of phenotypic variance with age in total cholesterol and apoB were almost entirely due to accumulation of unique environmental experiences in life. No consistent age trends were found for

triglycerides and apoA1. We found a similar pattern for apoB, with an increase in total variance after menopause that was due to an increase of unique environmental influences. For all other variables, with the exception of Lp(a), we observed increases in genetic variance after menopause, with a concomitant reduction in shared environmental variance, an increase in total variance, or both. For Lp(a), the best fitting model was identical in premenopausal and postmenopausal women, with high estimates of h² (87%). This finding is in accordance with evidence that ≈90% of the variation in Lp(a) is determined by a single gene, the apo(a) gene,^{28,29} whose influence is likely to be constant in life¹⁰ and is already fully expressed before the age of 1 year.³⁰

The limited number of longitudinal studies available suggests the existence of age-dependent gene expression in at least some of the lipids and lipoproteins. New genes are expressed during adolescence¹³ and during middle age in women,¹² and gene expression is different in childhood and adulthood.¹⁰ We found no evidence for different genes affecting lipids and (apo)lipoproteins before and after menopause. However, the detection of these effects in a cross-sectional twin design relies heavily on the number of informative twin pairs, ie, pairs discordant for their menopausal status. Although the number of DZ pairs discordant for menopause was reasonable, the number of MZ-discordant pairs was very small, probably because the age of menopause onset is itself under genetic control.¹⁷ Ideally, to exclude modest degrees of reduced genetic sharing, these results should be confirmed by a large longitudinal twin study that follows women through menopause, because hormonal and lipid levels change only gradually during the perimenopausal years. However, such a longitudinal design may never be practicable.

MZ pairs were, on average, 3.5 years older than DZ pairs, and a larger proportion was consequently postmenopausal. Furthermore, environmental variables such as fasting or HRT use are known to potentially have an influence on lipids and apolipoproteins. However, these factors are unlikely to have biased variance components estimates, because lipid values were adjusted for age, menopause, fasting status, and HRT use before model fitting analysis.

Although the generalizability of twin studies is sometimes disputed, the reported results are likely to be representative of singletons in the general population. We have recently shown that twins from the St. Thomas' UK Adult Twin Registry are similar to a population-based sample of >1000 women participating in the Chingford cohort study,¹⁵ London, involving a large number of health-related and cardiovascular variables.

Several practical implications follow from our findings. The importance of shared environmental influences on lipids and (apo)lipoproteins during early adulthood, as shown in the present study, may point to the likely success of family-based lifestyle interventions. Diet will most likely be an important component of such an intervention, because a recent study in young adult twins demonstrated that the association between diet on one hand and total cholesterol, LDL, and HDL on the other is due to environmental factors.³¹ The fact that no evidence was found for different genes influencing lipids before and after menopause implies that genome scan studies aiming to detect quantitative trait loci for lipids and (apo)lipoproteins can pool data from premenopausal and postmenopausal women. However, they may want to focus on the latter group, in which genetic variance seems to be larger. Future studies will need to determine the specific genes responsible for the increase in genetic variance after menopause. Polymorphisms underlying LDL receptor function and hepatic lipase activity are likely candidates because they are influenced by estrogen and progestin action.²

In conclusion, we have demonstrated that genetic influences on lipid and (apo)lipoprotein levels, except for apoB and Lp(a), are larger after menopause, whereas at the same time, shared environmental influences tend to be smaller or even disappear. No evidence was found that lipids are influenced by different genes before and after menopause. The importance of shared environmental influences on lipids and (apo)lipoproteins during early adulthood points to the potential for family-based lifestyle interventions. Human genetic studies aiming to optimize power for detection of quantitative trait loci underlying the genetic lipid variance, on the other hand, may want to focus on older individuals in which the genetic variance seems to be larger.

Acknowledgments

H. Snieder was sponsored by the British Heart Foundation (FS/99050). The St. Thomas' Twin Registry receives further funding from the Arthritis and Rheumatism Campaign, the Wellcome Trust, the Chronic Disease Research Foundation, and Gemini Genomics/Sequenom Inc. We wish to thank the research nurses for skillful data collection and all our twin volunteers for their support.

References

- Gordon T, Kannel WB, Hjortland MC, McNamara PM. Menopause and coronary heart disease: the Framingham Study. *Ann Intern Med.* 1978; 89:157–161.
- Bruckert E, Turpin G. Estrogens and progestins in postmenopausal women: influence on lipid parameters and cardiovascular risk. *Horm Res.* 1995;40: 305–311.
- Fukami K, Koike K, Hirota K, Yoshikawa H, Miyake A. Perimenopausal changes in serum lipids and lipoproteins: a 7-year longitudinal study. *Maturitas.* 1995;22:193–197.
- Schaefer EJ, Famon-Fava S, Cohn SD, Schaefer MM, Ordovas JM, Castelli WP, Wilson PW. Effects of age, gender, menopausal status on plasma low density lipoprotein cholesterol and apolipoprotein B levels in Framingham Offspring Study. *J Lipid Res.* 1994;35:779–792.
- Kuller LH, Meilahn EN, Cauley JA, Gutai JP, Matthews KA. Epidemiologic studies of menopause: changes in risk factors and disease. *Exp Gerontol.* 1994;29:495–509.
- McKinlay SM. The normal menopause transition: an overview. *Maturitas.* 1996;23:137–145.
- Rifai N, Chapman JF, Silverman LM, Gwynnes JT. Review of serum lipids and apolipoproteins in risk-assessment of coronary heart disease. *Ann Clin Lab Sci.* 1988;18:429–439.
- Reilly L, Kottke BA, Sing CF. The effects of generation and gender on the joint distributions of lipid and apolipoprotein phenotypes in the population at large. *J Clin Invest.* 1990;43:921–940.
- Boomsma DI, Kempen HM, Gevers LJ, Havekes L, de Knijff P, Frants RR. Genetic analysis of sex and generation differences in plasma lipid lipoprotein and apolipoprotein levels in adolescent twins and their parents. *Genet Epidemiol.* 1996;13:49–60.
- Snieder H, Doornen LJP, Boomsma DI. The age dependency of gene expression for plasma lipids, lipoproteins, and apolipoproteins. *Am J Hum Genet.* 1997;60:638–650.
- Snieder H, van Doornen LJP, Boomsma DI. Dissecting the genetic architecture of lipids, lipoproteins, and apolipoprotein: lessons from twin studies. *Arterioscler Thromb Vasc Biol.* 1999;19:2826–2834.
- Friedlander Y, Austin MA, Newman MA, Edwards K, Mayer-Davis EJ, King M-C. Heritability of longitudinal changes in coronary-heart-disease risk factors in women twins. *Am J Hum Genet.* 1997;60:1502–1512.
- Nance WE, Bodurtha JN, Eaves LJ, Hewitt J, Maes H, Segrest J, Meyer J, Neale MC, Schieken R. Models for the longitudinal genetic analysis of same-age twins: application to HDL cholesterol. *Twin Res.* 1998;1:3–8.
- Harris EL, Falk RT, Goldstein AM, Park LP. Clustering of high density lipoprotein cholesterol levels in premenopausal and postmenopausal female twins. *Genet Epidemiol.* 1993;10:563–567.
- Andrew T, Hart D, Snieder H, de Lange M, Spector TD, MacGregor AJ. Are twins and singletons comparable? A study of disease-related and lifestyle characteristics in adult women. *Twin Res.* 2001;4:464–477.
- Spector TD, Cicuttini FM, Baker JR, Loughlin J, Hart D. Genetic influences on osteoarthritis in women: a twin study. *BMJ.* 1996;312:940–944.
- Snieder H, MacGregor AJ, Spector TD. Genes control the cession of a woman's reproductive life: a twin study of hysterectomy and age at menopause. *J Clin Endocrinol Metab.* 1998;83:1875–1880.
- Hunter D, De Lange ME, Andrew T, Snieder H, MacGregor AJ, Spector TD. Genetic variation in bone mineral density (BMD) and calcaneal ultrasound: a study of the influence of menopause using female twins. *Osteoporos Int.* 2001;12:406–411.
- Friedewald WT, Levy RT, Fredrickson DS. Estimation of the concentration of low density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. *Clin Chem.* 1972;18:499–502.
- Neale MC, Cardon LR. *Methodology for Genetic Studies of Twins and Families.* Dordrecht, the Netherlands: Kluwer Academic Publishers; 1992.
- STATA Corp. *STATA Statistical Software, Release 5.0.* College Station, Tex: STATA Corp; 1997.
- MacGregor AJ. Practical approaches to account for bias and confounding in twin data. In: Spector TD, Snieder H, MacGregor AJ, eds. *Advances in Twin and Sib-Pair Analysis.* London, UK: Greenwich Medical Media; 2000: 35–52.
- Snieder H. Path analysis of age-related disease traits. In: Spector TD, Snieder H, MacGregor AJ, eds. *Advances in Twin and Sib-Pair Analysis.* London, UK: Greenwich Medical Media; 2000:119–129.
- Lange K, Westlake J, Spence M. Extensions to pedigree analysis, III: variance components by the scoring method. *Ann Hum Genet.* 1976;39: 485–491.
- Neale MC, Boker SM, Xie G, Maes HH. *Mx: Statistical Modeling.* Richmond, Va: Department of Psychiatry, Virginia Commonwealth University; 1999.
- Hopper JL. Why "common" environmental effects" are so uncommon in the literature. In: Spector TD, Snieder H, MacGregor AJ, eds. *Advances in Twin and Sib-Pair Analysis.* London, UK: Greenwich Medical Media Ltd; 2000: 151–165.
- Iliadou A, Lichtenstein P, De Faire U, Pedersen NL. Variation in genetic and environmental influences in serum lipid and apolipoprotein levels across the lifespan in Swedish male and female twins. *Am J Med Genet.* 2001;102: 48–58.
- Boerwinkle E, Leffert CC, Lin J, Lackner C, Chiesa G, Hobbs HH. Apolipoprotein(a) gene accounts for greater than 90% of the variation in plasma lipoprotein(a) concentrations. *J Clin Invest.* 1992;90:52–60.
- DeMeester CA, Bu X, Gray RJ, Lusis AJ, Rotter JJ. Genetic variation in lipoprotein(a) levels in families enriched for coronary artery disease is determined almost entirely by the apolipoprotein(a) gene locus. *Am J Hum Genet.* 1995;56:287–293.
- Wang XL, Wilcken DEL, Dudman NPB. Early expression of the apolipoprotein(a) gene: relationships between infants' and their parents' serum apolipoprotein(a) levels. *Pediatrics.* 1992;89:401–406.
- McCaffery JM, Pogue-Geile MF, Muldoon MF, Debski TT, Wing RR, Manuck SB. The nature of the association between diet and serum lipids in the community: a twin study. *Health Psychol.* 2001;20:341–350.